

A new structural approach to study lipid-protein interactions within a viral envelope

Larisa Kordyukova

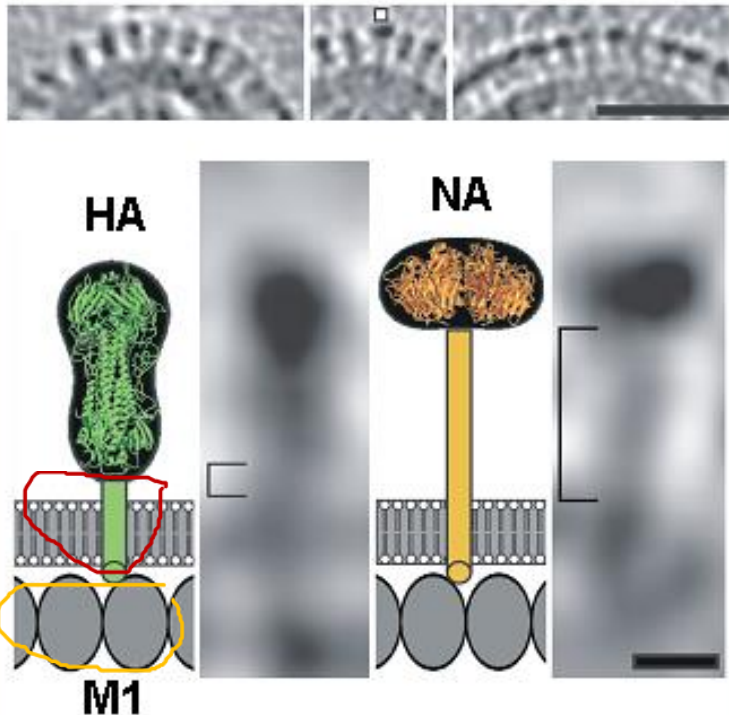
*Belozersky Institute of Physico-Chemical Biology,
Lomonosov Moscow State University*



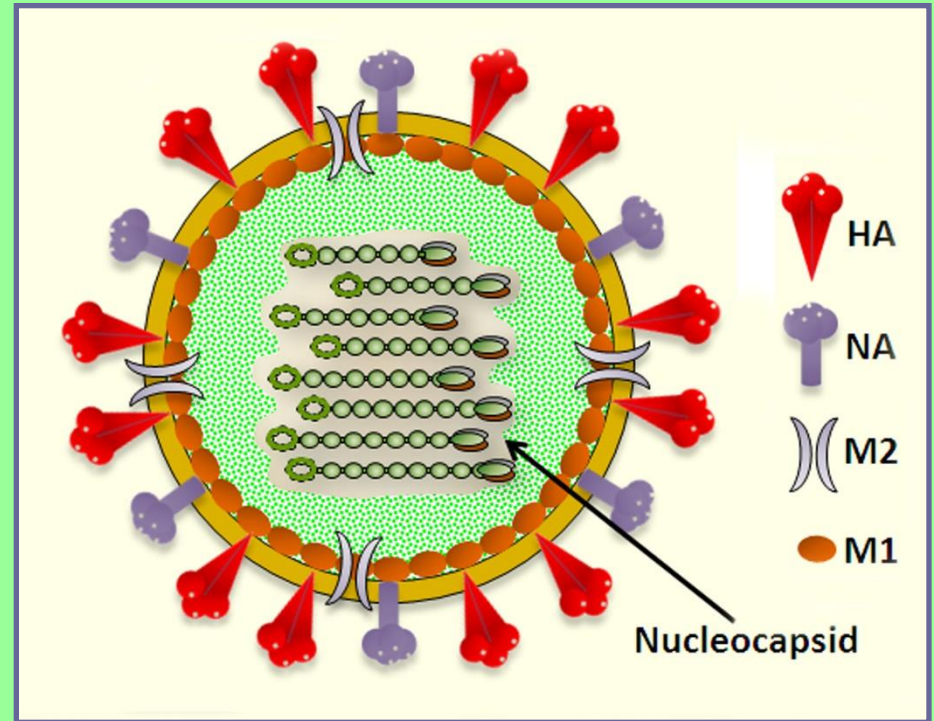
Orthomyxoviridae. Genus Influenzavirus A

HA:
Trimer

NA:
Tetramer



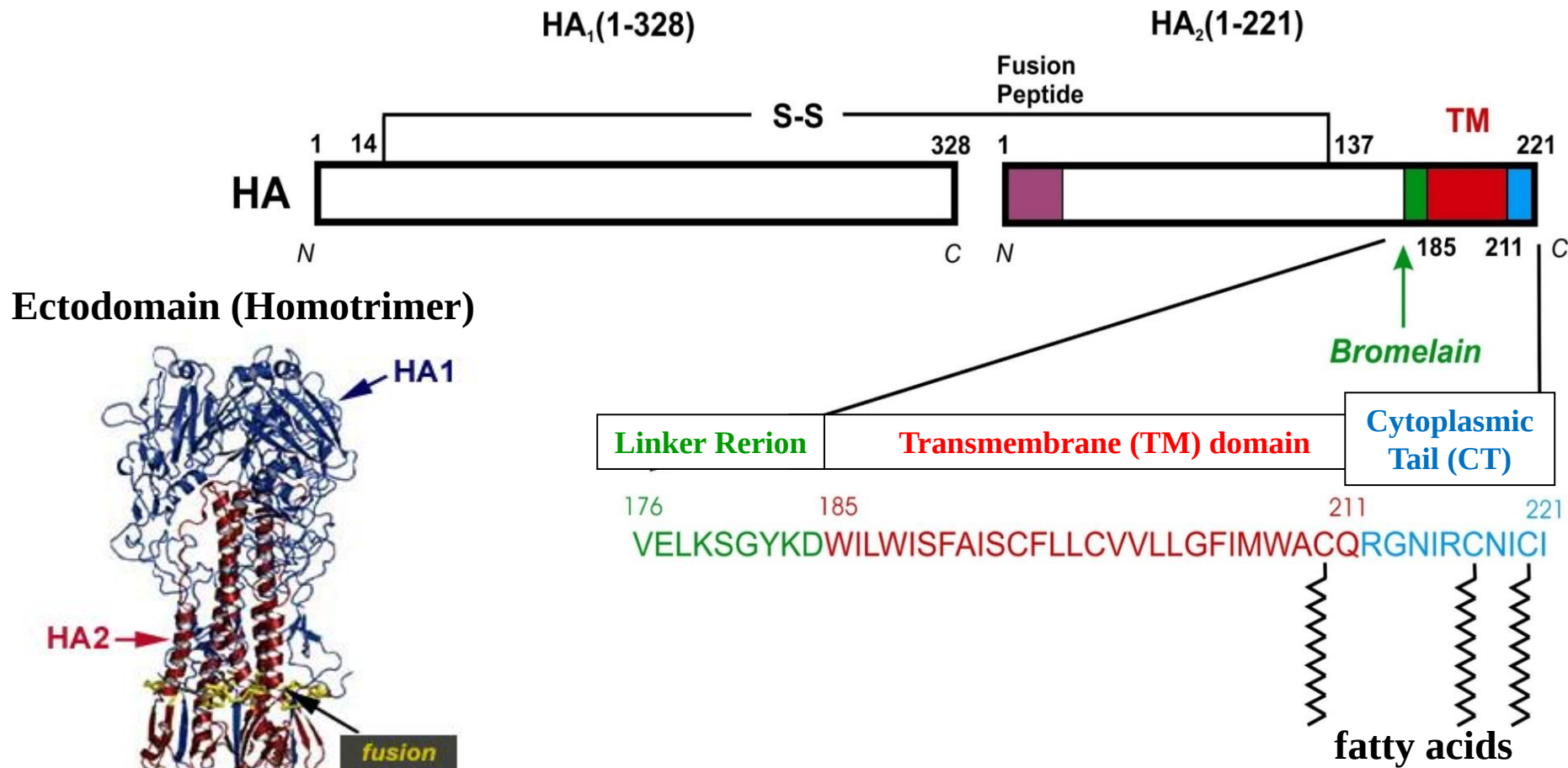
Harris et al., *PNAS*, 2006, 103: 19123



- Mechanisms of HA and NA interactions with a layer of M1 matrix lipid membrane are poorly understood
- Such interactions are critical for virus particles assembly and budding

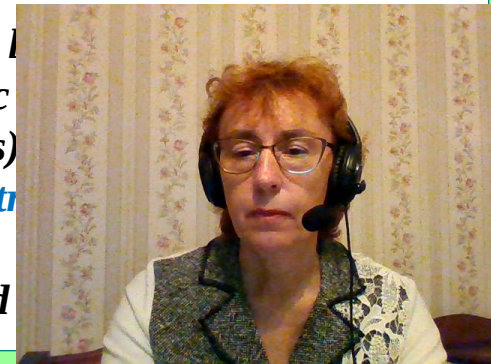


Structure of influenza hemagglutinin (HA)

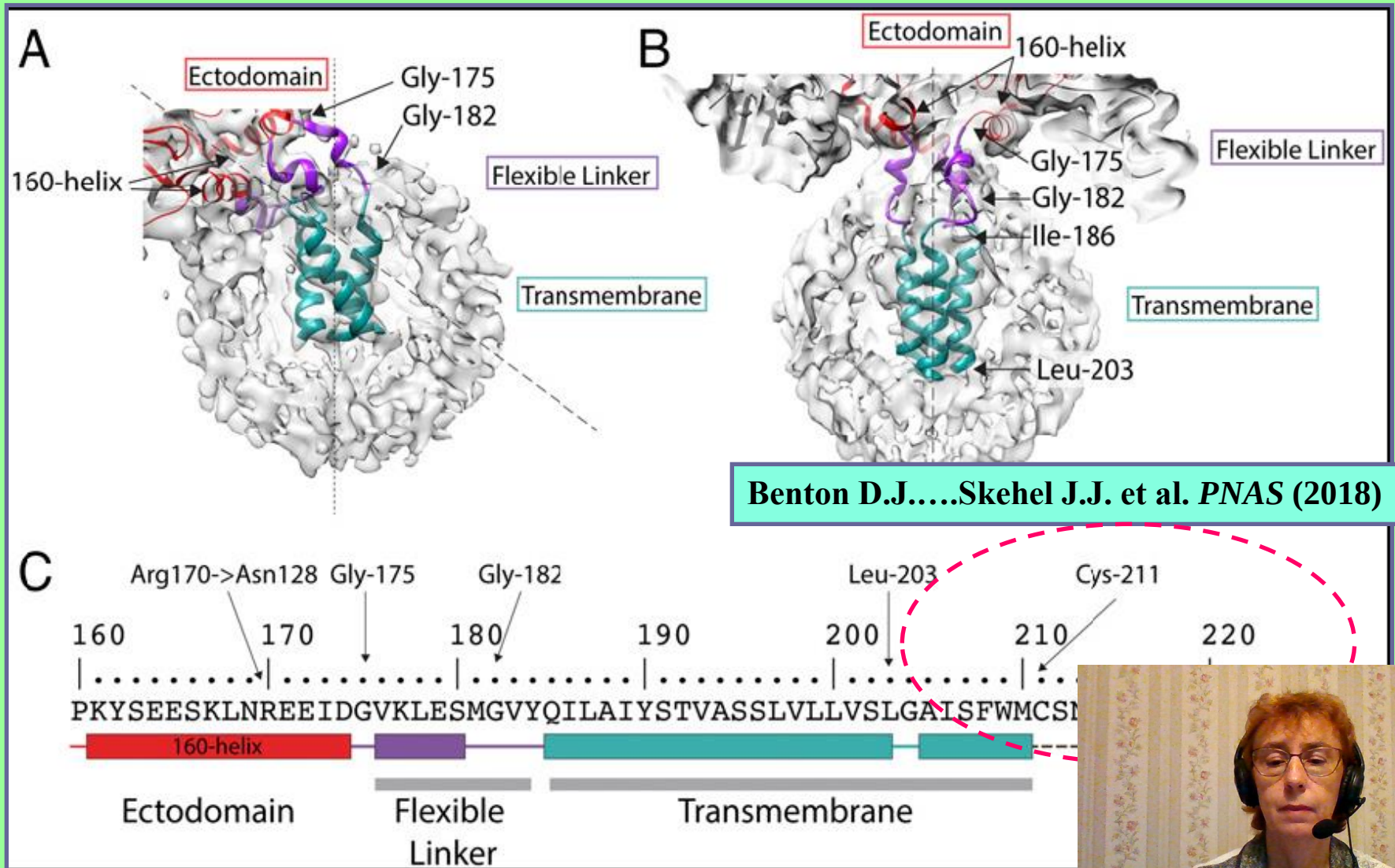


Anchoring segment

- HA monomer contains HA1 and HA2 chains
- X-ray data are available for the homotrimeric
- Anchoring segment (~45 amino acids)
- **transmembrane domain** + **cytoplasmic tail** (intracellular)
- Several cysteine residues in C-terminus modified with long saturated fatty acids bound

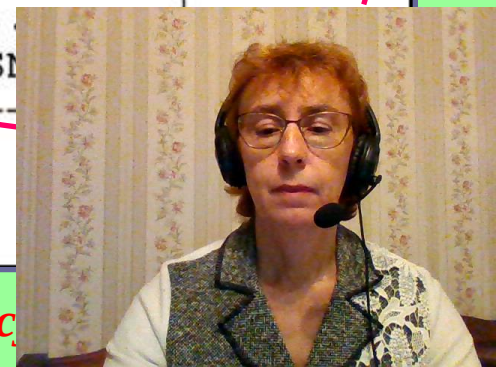


Cryo-EM: the N-terminal part of HA membrane anchor is resolved



Benton D.J.....Skehel J.J. et al. *PNAS* (2018)

BUT: The C-terminal part of TMD and the CT including S-acylated c



MALDI-TOF MS to study S-acylation of HA

A/chicken/Germany/N/49 (H10N7)

HA 520-561 +

3 Pal
2 Pal / 1 Str
1 Pal / 2 Str

HA 518-561 +

3 Pal
2 Pal / 1 Str
1 Pal / 2 Str

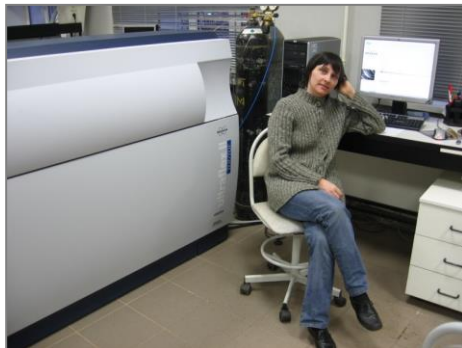
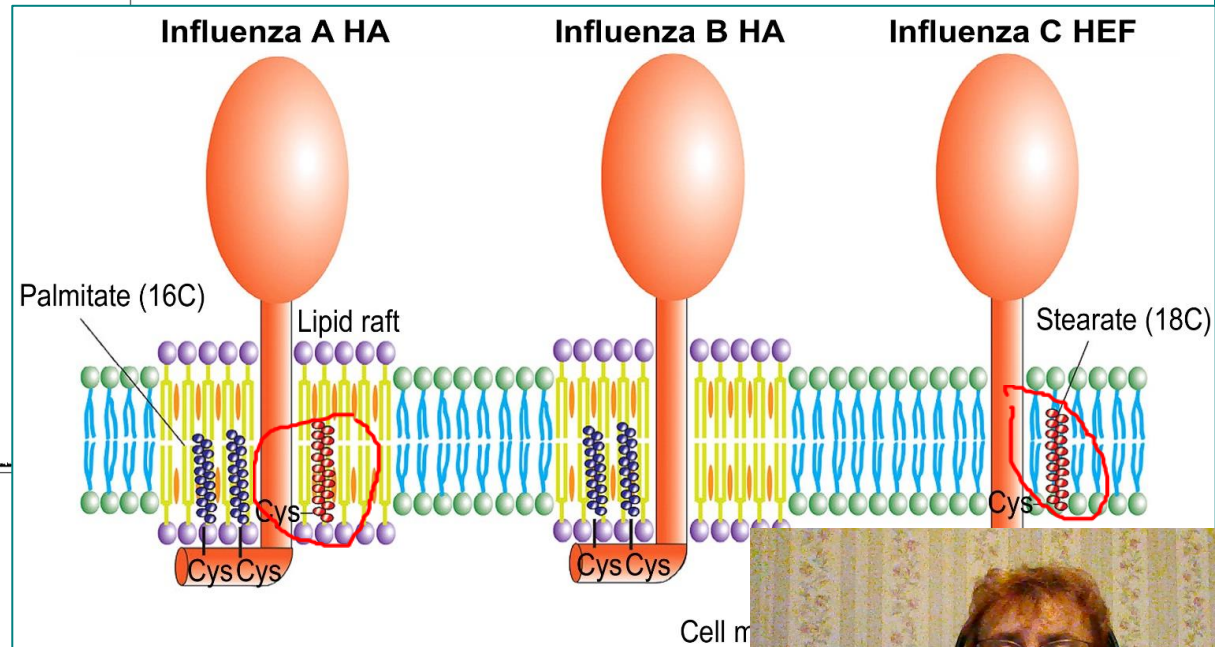
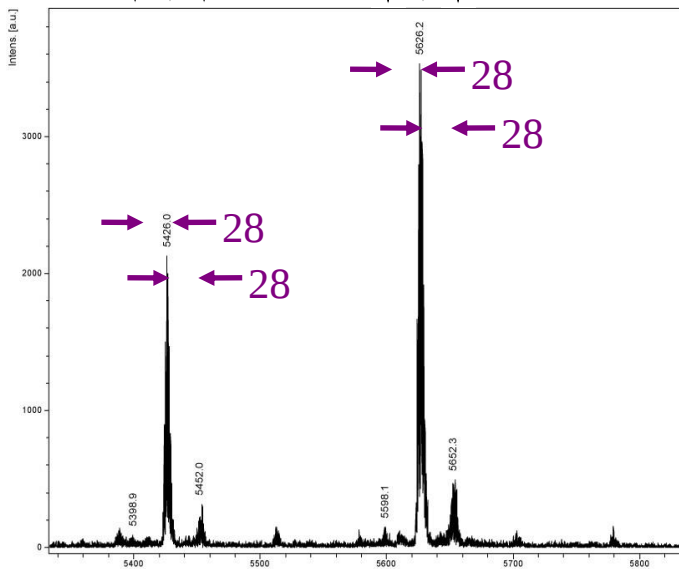
Palmitate (C16:0) = +238 m/z

Stearate (C18:0) = +266 m/z

266-238=28 m/z

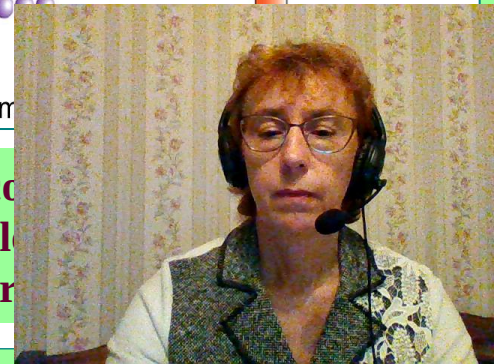


PD Dr. Michael Veit
Free University Berlin



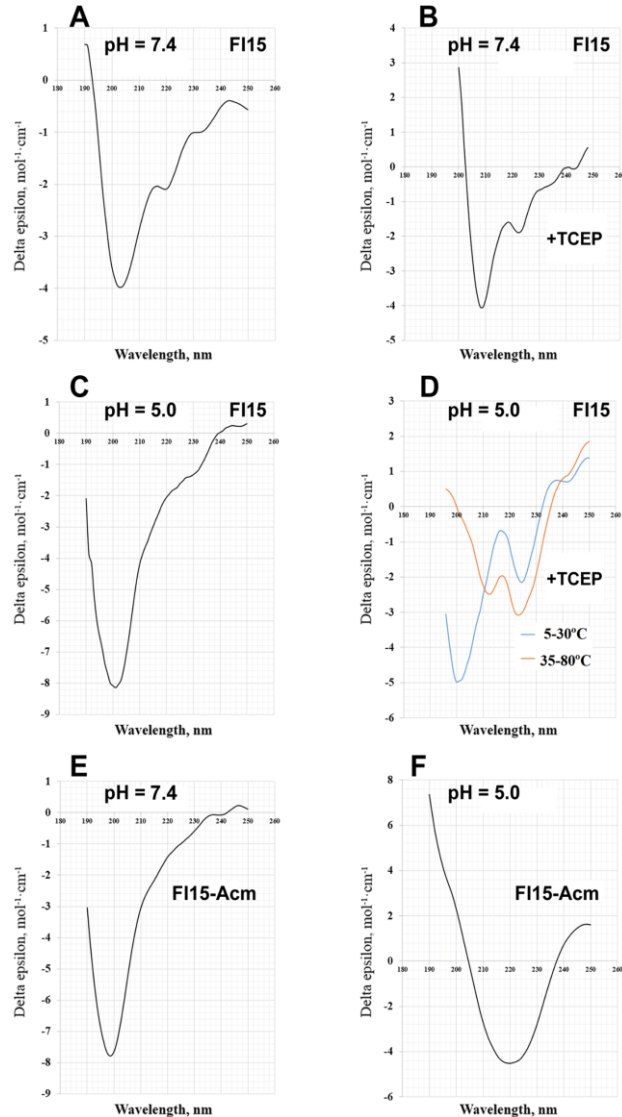
Dr. Marina Serebryakova
Lomonosov Moscow State University

Glyco
envel
differ

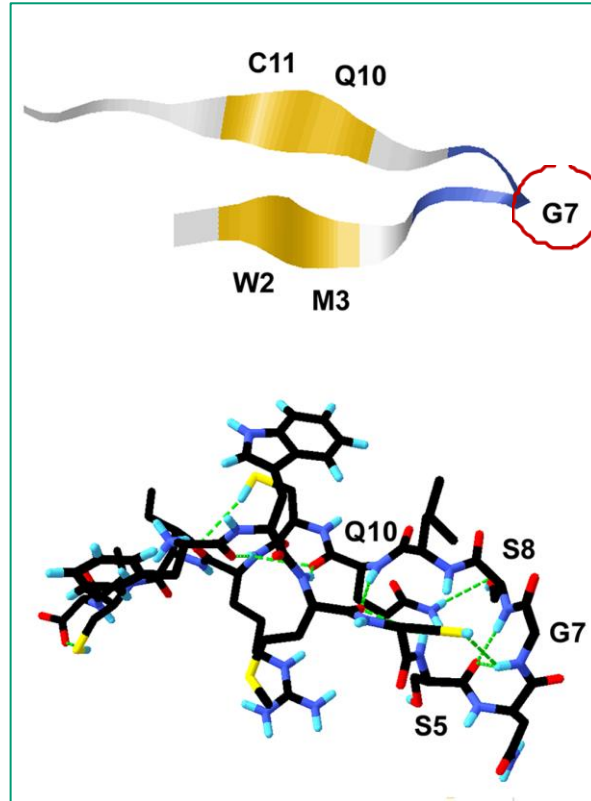


Cytoplasmic tail of HA is β -structural

Circular dichroism analysis



Molecular modeling



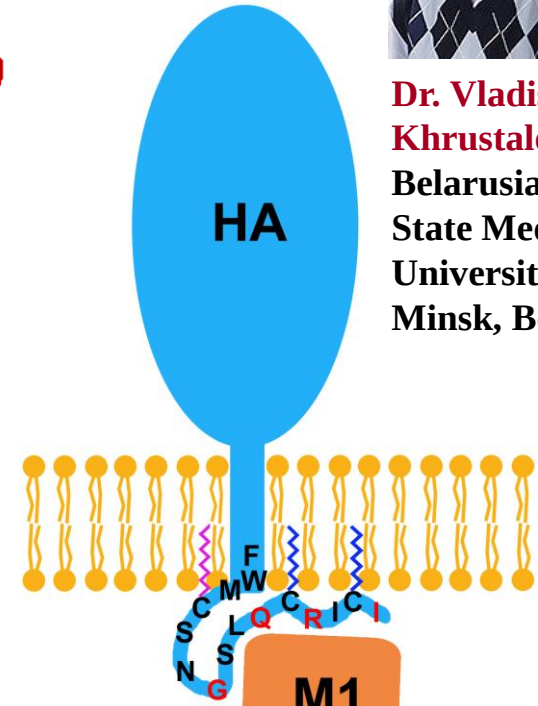
Homology-based 3D models of the FI15 peptide

FI15: $\text{NH}_2\text{-FWMC SNGSLQCRICI-COOH}$

FI15-Acm: $\text{NH}_2\text{-FWMC* SNGSLQC*RI}$
 *acetaminomethylated -SH groups of cy

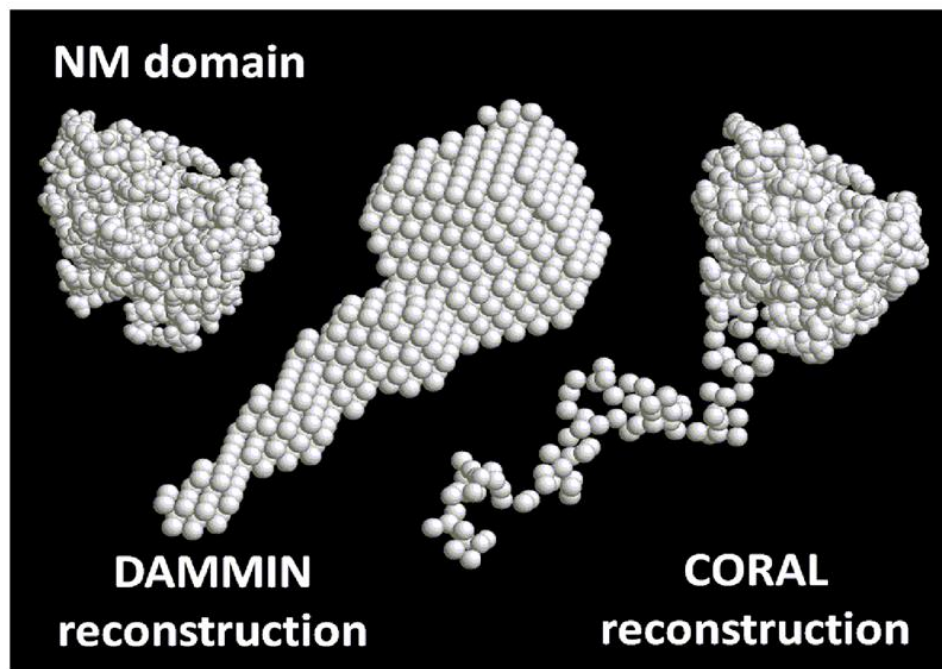


Dr. Vladislav V. Khrustalev
 Belarusian State Medical University,
 Minsk, Belarus



Small Angular X-ray Scattering (SAXS) to analyze protein structure in solution

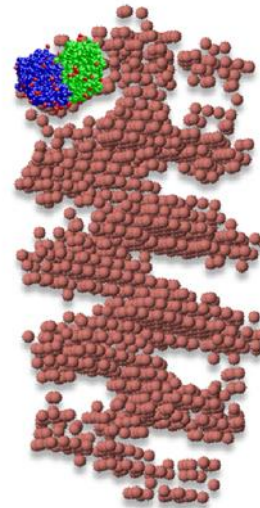
SAXS analysis:
full-length M1 model with globular
NM-domain and flexible C-domain



Shtykova E.V. et al. PLoS One (2013) 8(12):e82431;
Shtykova E.V. et al. Sci. Rep. (2017) 7(1):16793.

**EMBL, Hamburg Outstation,
“Deutsche Synchrotrone” (DESY), Germany**

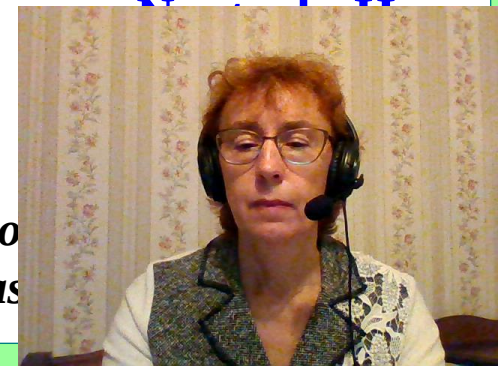
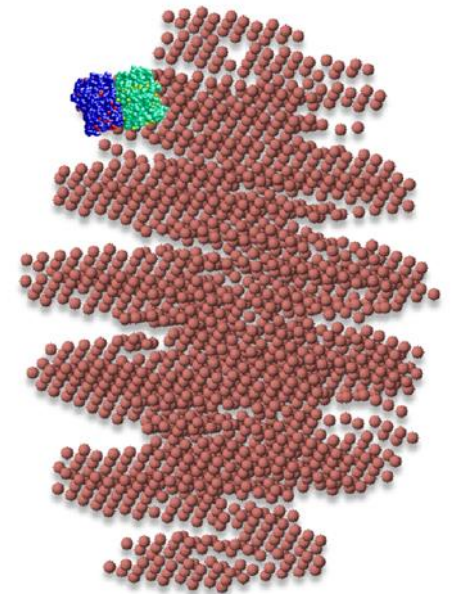
**Small
cluster**



Acidic pH
NM-domain
1AA7 PDB

M1-M1 association
(association increases)

**Large
cluster**



SAXS to model M1-lipid and M1-HA interactions?

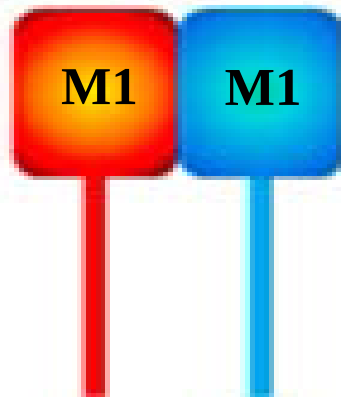
Transmembrane domain

- *M1 - negatively charged (anionic) lipids*
- *M1 - liquid-ordered (raft) lipids*
- *M1 - hemagglutinin cytoplasmic tail*

Stear PalPal

Viral Membrane

Cytoplasmic tail



Liposomes:

Extrusion through 100 nm pore

- (1) Synthetic - with anionic lipids
- (2) Synthetic - with anionic lipids
- (3) Native (viral) lipids (4)
- (4) Native (viral) lipids &

SAXS analysis
of lipid vesicles
mixtures



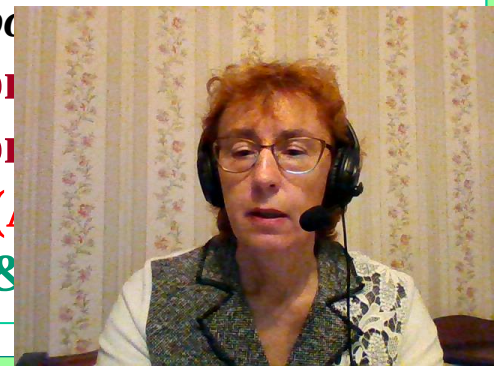
M1

from A/PR8/8/34

(acid solubilization)

Adjusted to pH = 6.8

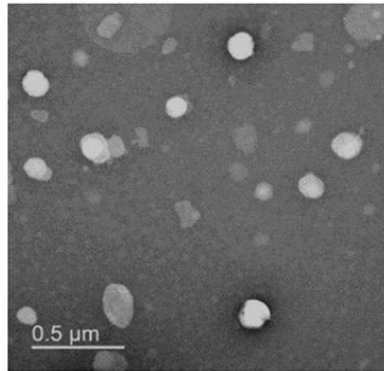
+



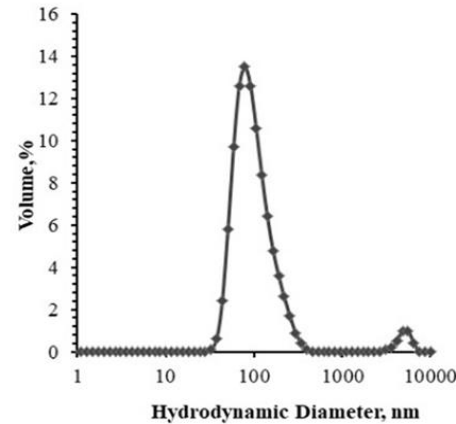
EM and DLS characterization of liposomes

Liposomes extruded through 100 nm pores

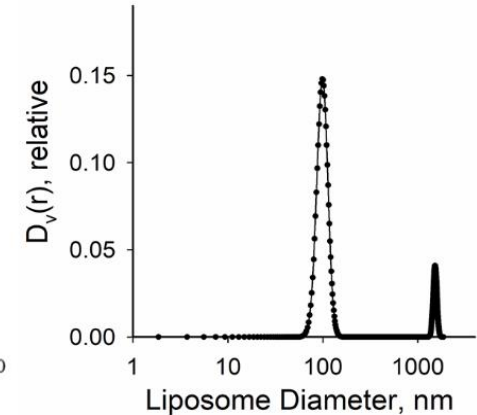
10 mol.% DOPS
90 mol.% DOPC



(a)

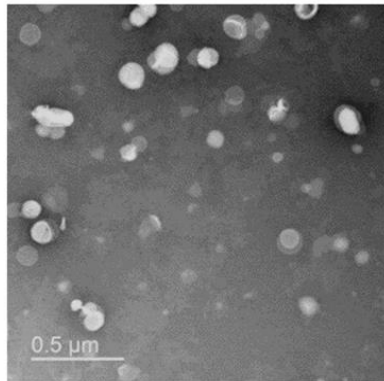


(b)

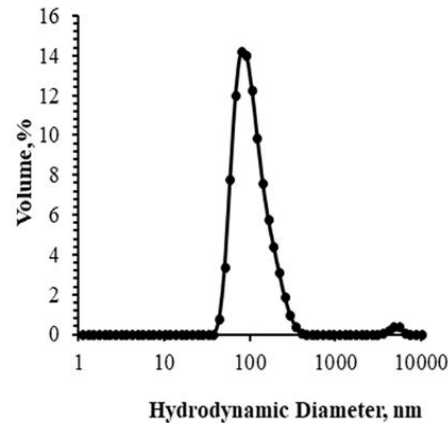


(c)

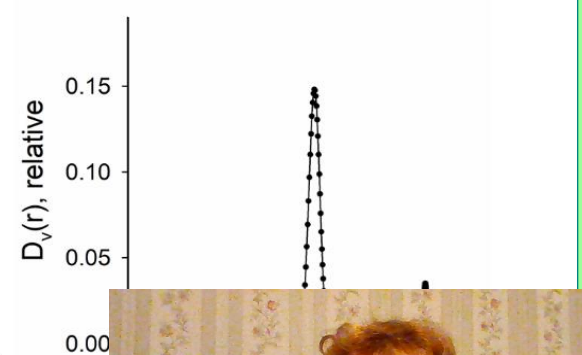
30 mol.% DOPS
70 mol.% DOPC



(d)

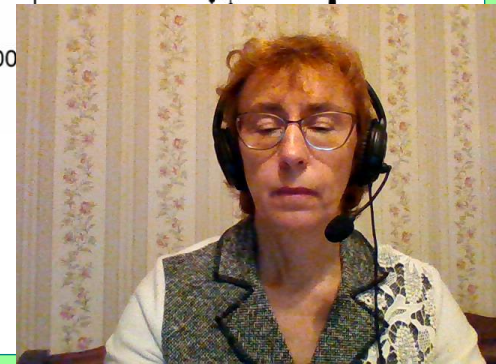


(e)

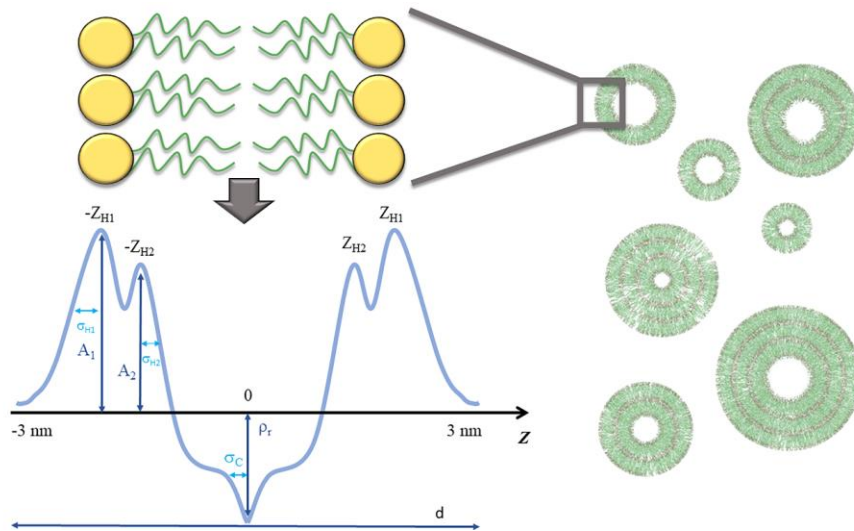


Electron Microscopy
(negative contrasting)

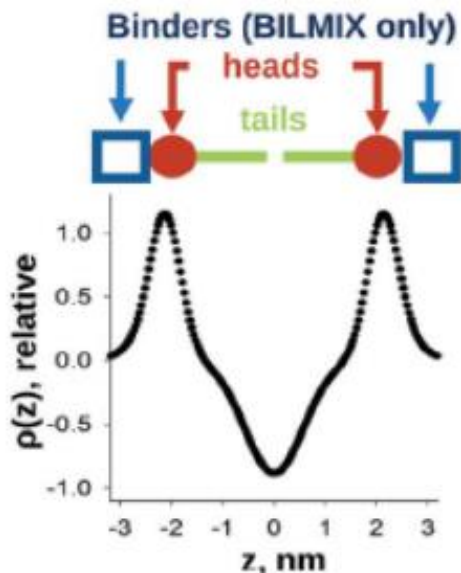
Dynamic Light Scattering



Mixtures of lipid vesicles: SAXS data analysis



Bilayer electron density



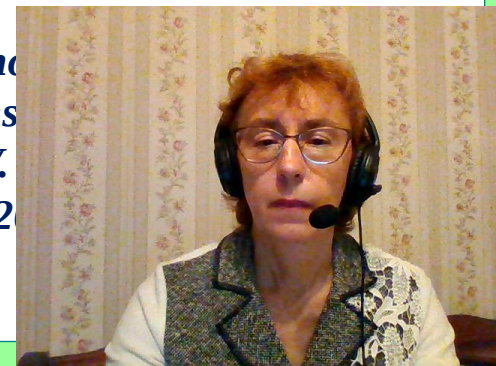
+ protein

Mixtures of lipid vesicles, SAXS analysis

Program BILMIX* (Bilayer Lipid MIXtures)
restores
the **electron density of a lipid bilayer** and
simultaneously generates the **size distribution**
of the unilamellar lipid vesicles
(using either spherical or ellipsoidal models)

BILMIX allows also the modelling of
asymmetric electron-density profiles,
e.g. **proteins** associated
with the inner or outer leaflets of the
liposome

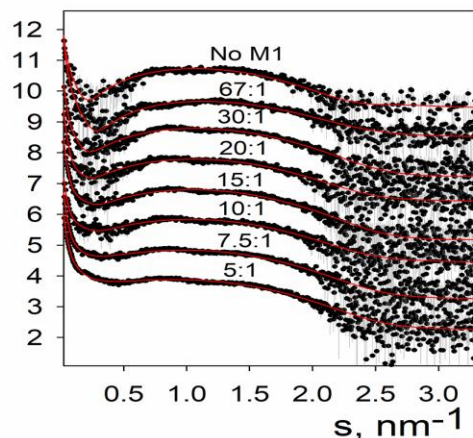
* Konarev P.V., Petoukhov
Fedorova N.V., Volynskiy
Batishchev O.V.
J. Appl. Cryst. (2004)



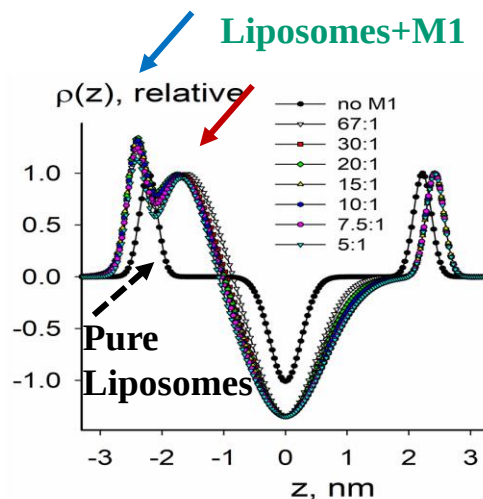
SAXS to model M1 - anionic lipid interactions

SAXS experimental curves and BILMIX calculations

Igl, relative



Restored electron density profiles

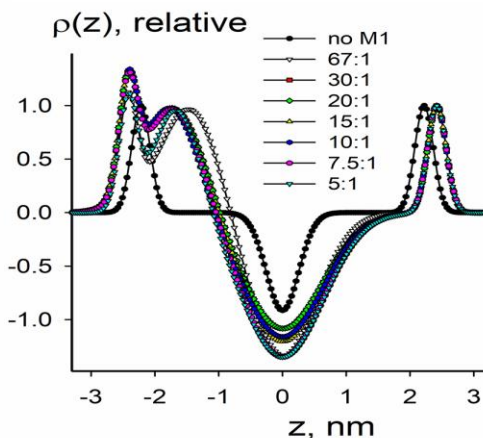
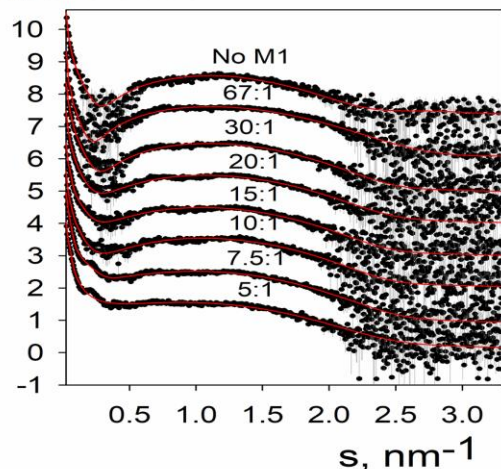


In two-component liposomes, the left maximum splits into two sub-peaks upon loading liposomes with M1: the left (M1-associated) sub-peak with increased intensity implies charged lipid condensation. The right sub-peak indicates outer monolayer bending inside



Igl, relative

10% DOPS + 90% DOPC



30% DOPS + 70% DOPC

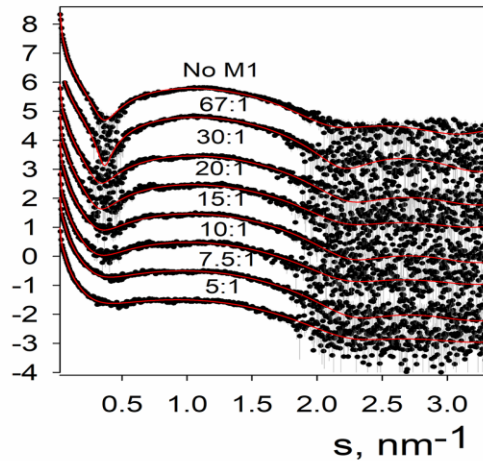
P12 beam
synchrotron
III (DESY)
ATSA
BILMIX



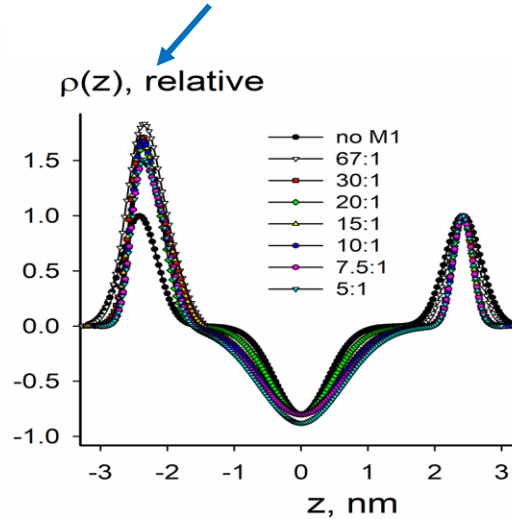
SAXS to model M1 – raft lipid interactions

SAXS experimental curves and BILMIX calculations

Igl, relative



Restored electron density profiles

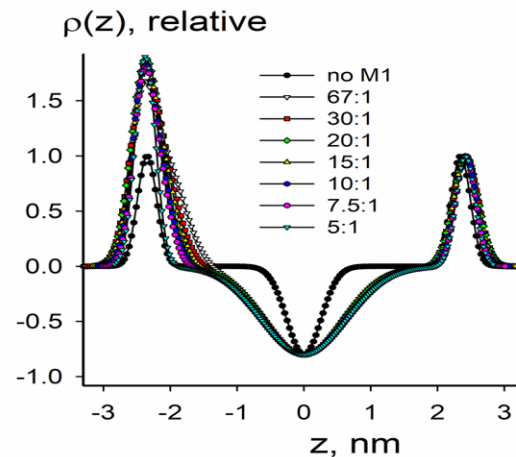
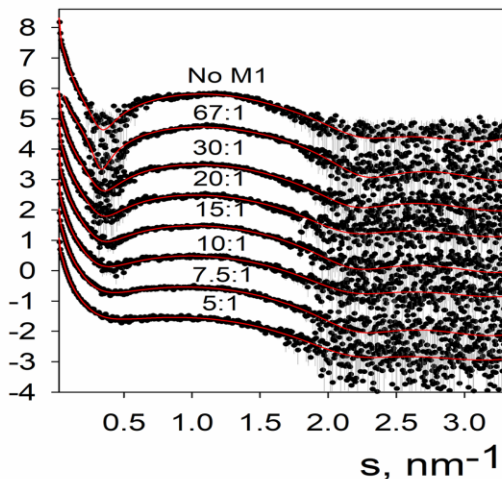


In four-component liposomes, we do not observe a separated protein-associated peak after loading with M1; the intensity of the left maximum increases. We may suggest that M1 introduces its helices into the lipid bilayer.

Hypothesis

- Adsorption of M1 protein at the liposomes containing **phosphatidylserine** leads to condensation of lipids underneath;
- Imbalance in the area of lipid monolayers occurs, leading to the bending of the membrane inside the of protein
- **Cholesterol** process a membran

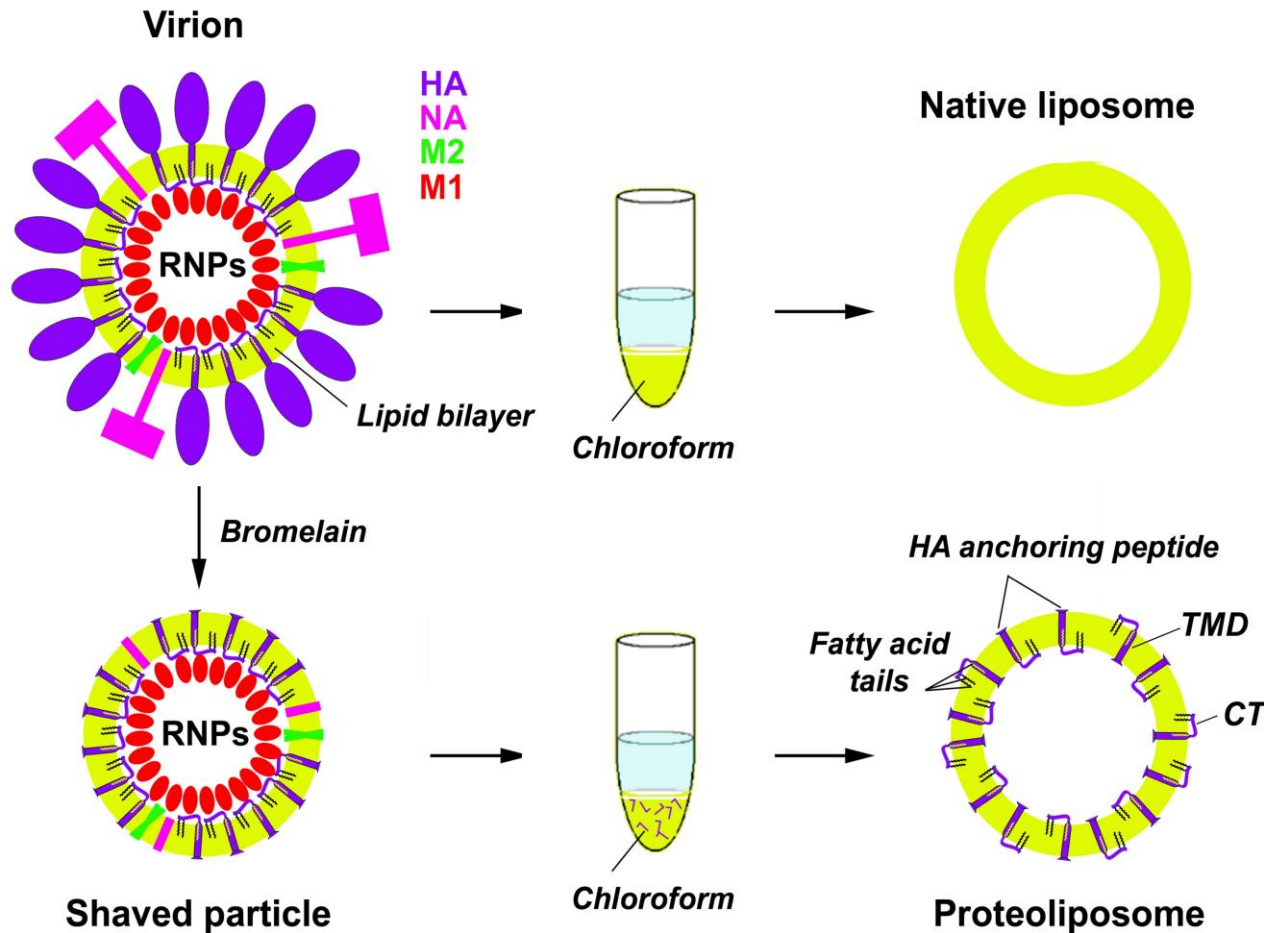
Igl, r **30% bPS + 10% POPC + 40% SM + 20% Chol**



20% bPS + 13.3% POPC + 33.3% SM + 33.3% Chol



Liposomes from viral lipids +/- HA LI45 peptide



Native liposomes
and proteoliposomes
prepared from
Influenza A/H1N1 virus

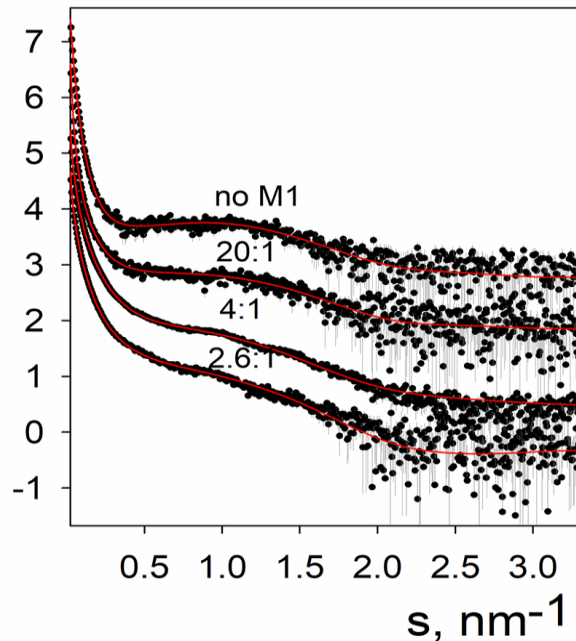
HA C-terminal peptide of H1N1 virus: "LI45" (45 amino acids)
NH₂-LESMGIYQILAIYSTVASSLVLLVSLGAISFWMCN¹GSIL²
triple palmitoylated at three conserved cysteine residues



SAXS to model M1 - viral lipid interactions

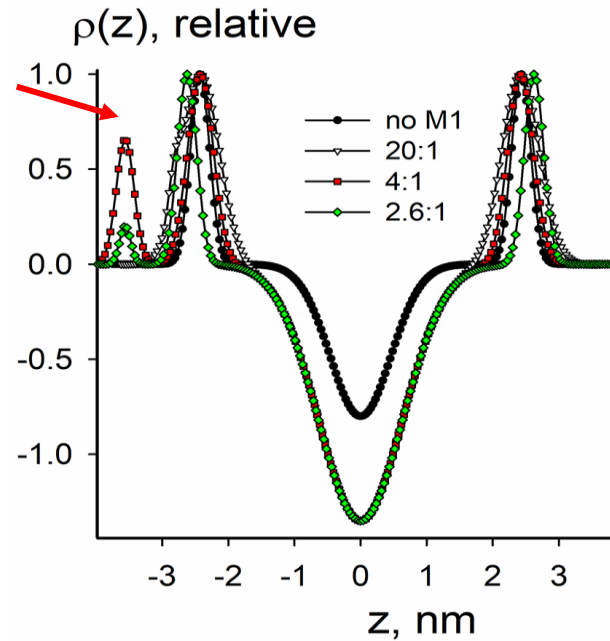
SAXS experimental curves and BILMIX calculations

$I(q)$, relative

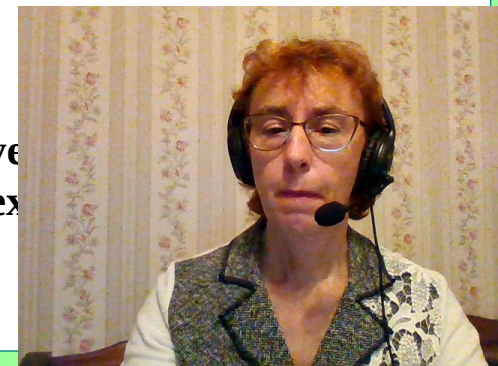


Restored electron density profiles

Native liposomes



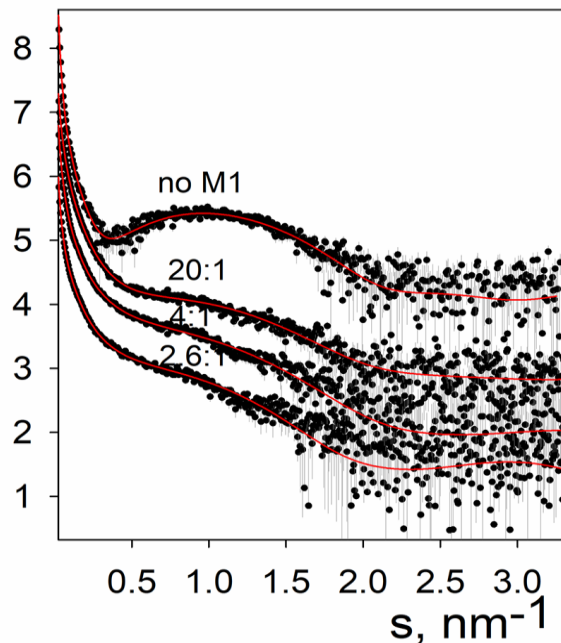
1. A drop of the electron density at the intermonolayer surface is observed upon adsorption of the M1 protein only (not in the case of pure liposomes);
2. The M1-associated peak appears that is completely separated from the second, lipid-associated peak;
3. We do not see shifting of the lipid-associated sub-peak closer to the bilayer center. Thus, membrane tubular invaginations are not formed in complex mixtures of viral lipids in contrast to synthetic charged vesicles.



SAXS to model M1 - HA interactions

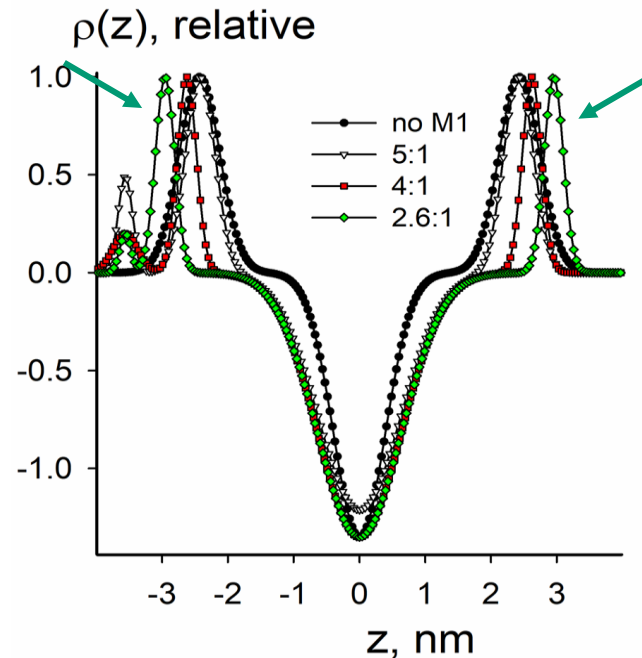
SAXS experimental curves and BILMIX calculations

$I(q)$, relative



Restored electron density profiles

Proteoliposomes



1. In contrast to native liposomes, a drop of the electron density at the intermonolayer surface **is observed even before loading with M1 protein.**
2. **This result indicates that triply palmitoylated HA LI45 peptides make lip bilayer more ordered, and viral membrane becomes raft-like;**
3. Two positive maxima move to the periphery from the bilayer centrum up adsorption with M1. **This effect is more pronounced in proteoliposomes compared to native liposomes: the whole width of their lipid bilayer increase by 2 nm (1 nm at each side).**



CONCLUSIONS

- M1 interaction with phosphatidylserine leads to condensation of the lipid in the protein-contacting monolayer thus resulting in formation of lipid tubules;
- This effect vanishes in the presence of the raft-forming constituents (sphingomyelin and cholesterol);
- Hemagglutinin anchoring peptides bearing three fatty acid residues demonstrate a specific role in ordering of viral lipid membrane into the raft-like one;
- Hemagglutinin anchoring peptides may stimulate the oligomerization of M1 on the lipid membrane to form a viral scaffold for subsequent budding of virion from the plasma membrane of the infected cell.



MAIN PARTICIPANTS:



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SAXS analysis



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M1 preparations & DLS measurements



Oleg Batishchev

Frumkin Institute of Physical Chemistry and Electrochemistry, Russian Academy of Sciences, Moscow, Russia

Biophysics of lipid membranes

Dmitri Svergun

P12 beamline (EMBL) of synchrotron storage ring Petra-III (DESY), Hamburg, Germany

Tatiana Timofeeva

D. I. Ivanovsky Institute of Virology, FSBI N. F. Gamaleya NRCM, Ministry of Health of Russian Federation, Moscow, Russia (**Virus**)

Sergei Abra

Department of Moscow State

Andrei Moi

Department of Moscow State



**Thank you for
your attention!**

